

Integrating vineyard pathogen control with sustainable nutrient and energy management



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Topics

- Application of BSM in pathogen control
 - Direct toxicity
 - Induction of systemic resistance



- Sustainable practices

- Compost application
- Anaerobic digestion
- Channelized aquatic scrubbers



Apple Replant Disease



Virgin



Replant

2nd leaf

Rootstocks vary in susceptibility to pathogens



+Rs

-Rs



+Rs

-Rs



Pathogens commonly responsible for apple replant disease:

- Fungi

- *Rhizoctonia solani*



- Oomycetes

- *Pythium* spp.



- Nematodes

- *Pratylenchus penetrans*



Soil amendment with Brassicaceae seed meals (BSM)



Disease control following BSM application

- **Biofumigation by isothiocyanates**
 - Important in control of nematodes and oomycetes
- **Enhancement of plant disease resistance**
 - BSM-induced shift in the soil microbial community

Release of ITC from BSM amended soil

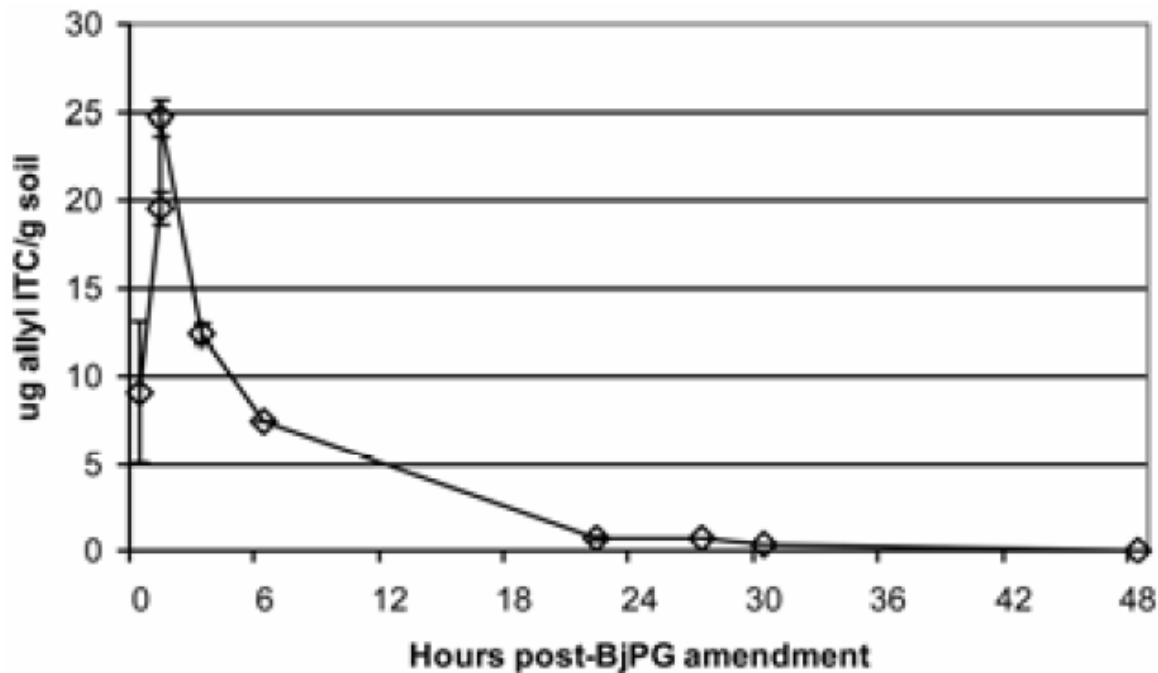
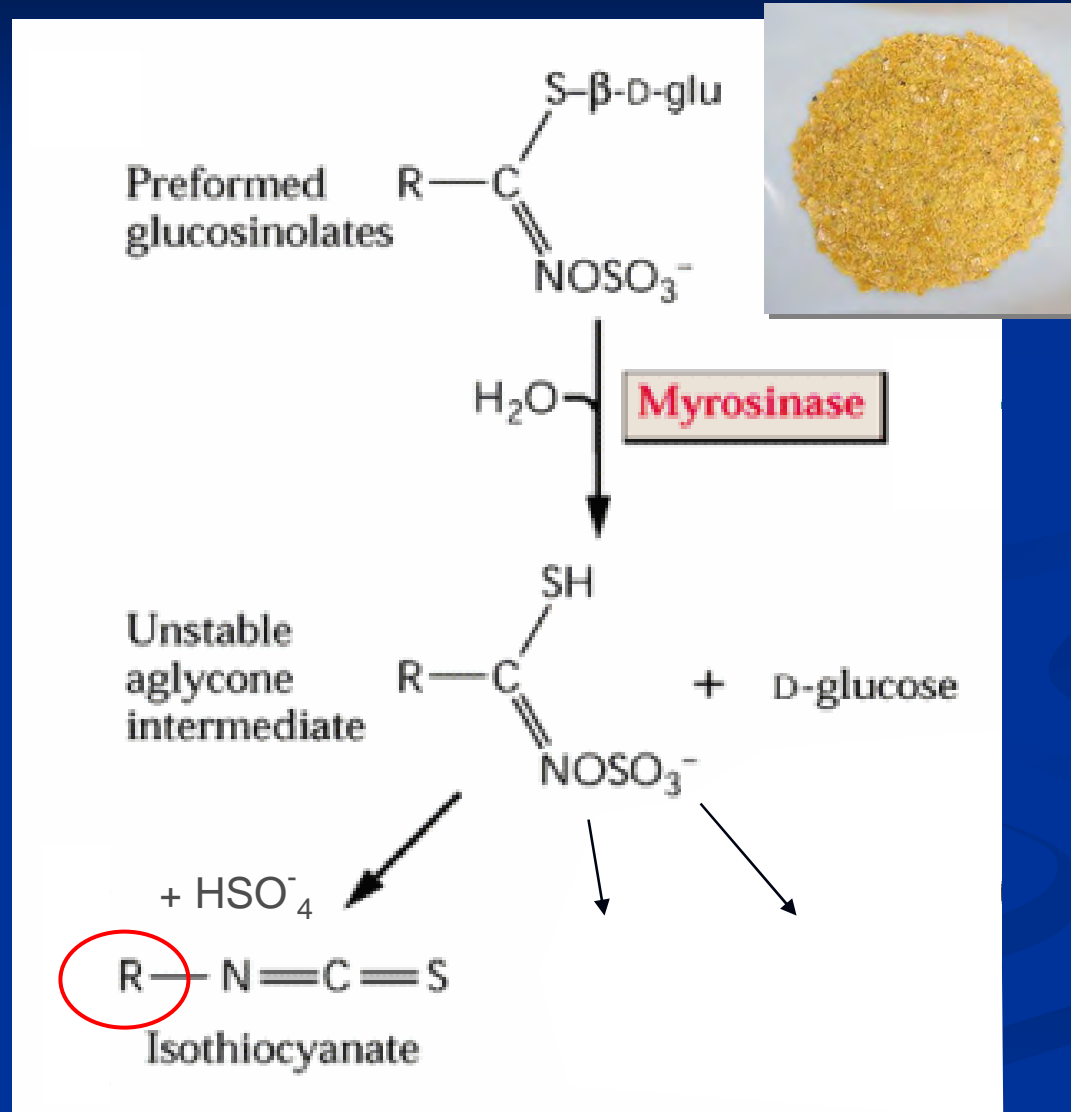


Fig. 1. Temporal pattern of allyl-isothiocyanate emission from Columbia View orchard soil amended with *Brassica juncea* cv. Pacific Gold seed meal (BjPG) as determined by monitoring concentration in the headspace of a chamber by gas chromatography. Seed meal was added to soil at a concentration of 0.5% (vol/vol). Bars = standard deviation of the mean.

ITC release from glucosinolate hydrolysis



ITC toxicity

- Type of ITC
- Concentration of ITC
- Nematode species

BSM glucosinolate composition varies

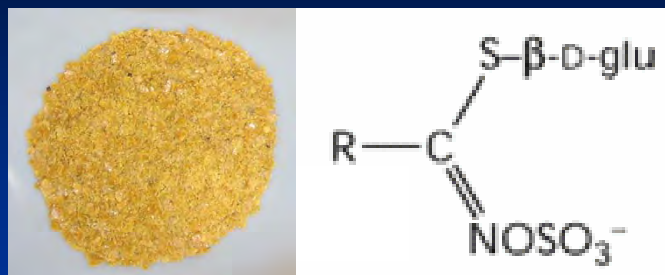
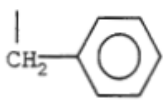
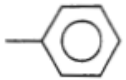
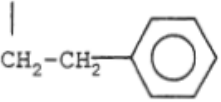


TABLE 1. Isothiocyanate, origin, structure, molecular weight, and common name of parent glucosinolate tested^a

Isothiocyanate	Plant species	Plant part	Structure of side chain R	Molecular weight	Glucosinolate common name
Allyl	<i>Armoracia lapathifolia</i> , <i>Brassica juncea</i> , <i>B. napus</i> , <i>B. oleraceae</i>	Seed, leaf, root, stem	$\text{CH}_2(\text{CH})_2\text{CH}_3$	99.2	Sinigrin
Benzyl	<i>Carica papaya</i> , <i>B. hirta</i> , <i>Lepidium sativum</i>	Seed, leaf, root, stem		149.2	Glucotropolin
Butyl	<i>A. lapathifolia</i> , <i>Capparis flexuosa</i>	Seed, leaf, root, stem	$\text{CH}_2(\text{CH})_2\text{CH}_3$	115.2	
Ethyl	<i>Lepidium menziesi</i>	Seed	CH_2CH_3	87.1	Glucolepdiin
Methyl	<i>Capparis</i> spp.	Seed	CH_3	73.1	Glucocapparin
Phenyl	<i>A. lapathifolia</i>			135.2	
4-Methylsulfinyl(butyl)	<i>B. oleraceae</i>	Seed, leaf, root, stem	$\text{CH}_2(\text{CH})_3\text{-S-CH}_3$	177.3	Glucoraphanin
2-Phenylethyl	<i>A. lapathifolia</i> , <i>B. juncea</i> , <i>B. napus</i> , <i>B. hirta</i>	Seed, leaf, root, stem		163.2	Gluconasturtiin

^a Data from Fahey et al. (11) and Brown and Morra (5).

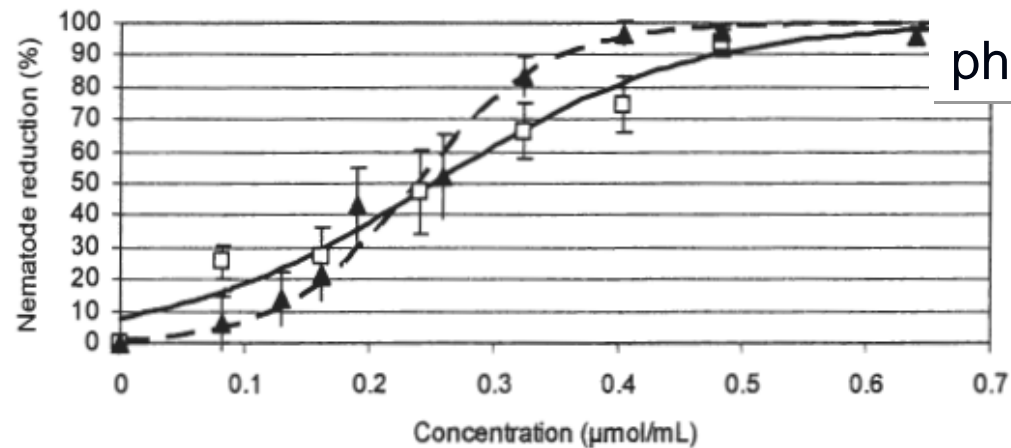
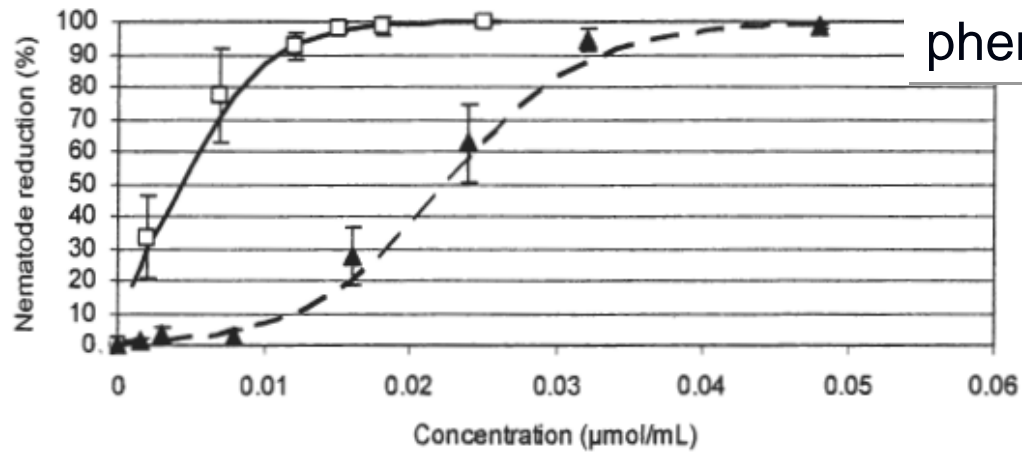
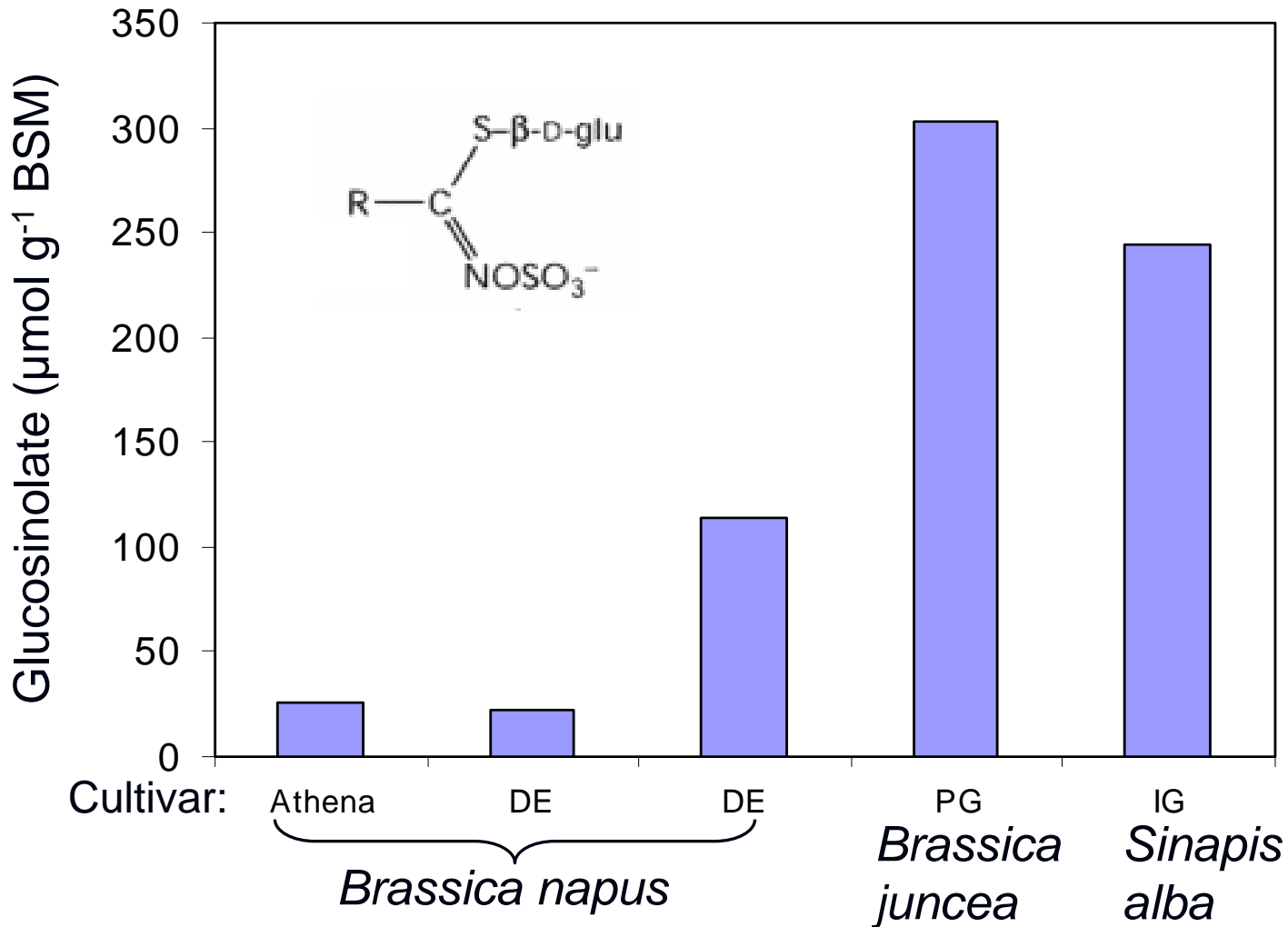
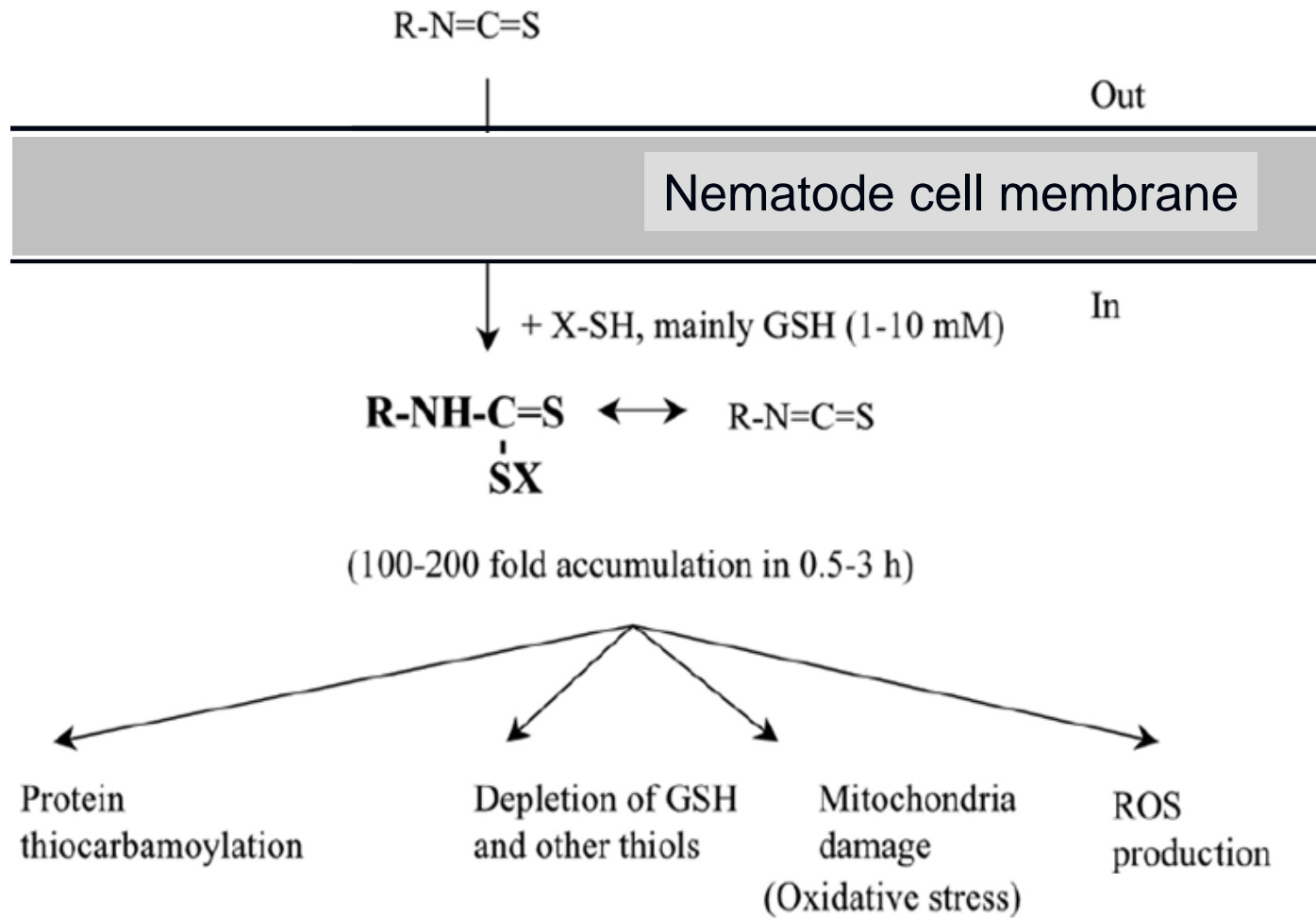


Fig. 2. Response curves for percent reduction of *Meloidogyne javanica* (▲) and *Tylenchulus semipenetrans* (□) exposed to concentration ranges of **A**, 2-phenylethyl and **B**, phenyl isothiocyanates. Vertical bars represent the 95% confidence interval for each mean.

BSM glucosinolate levels vary



Cellular effects of ITCs



BSM application in nematode control: Apple replant disease



Two mechanisms of initial BSM-induced nematode suppression



glucosinolate \rightarrow ITC

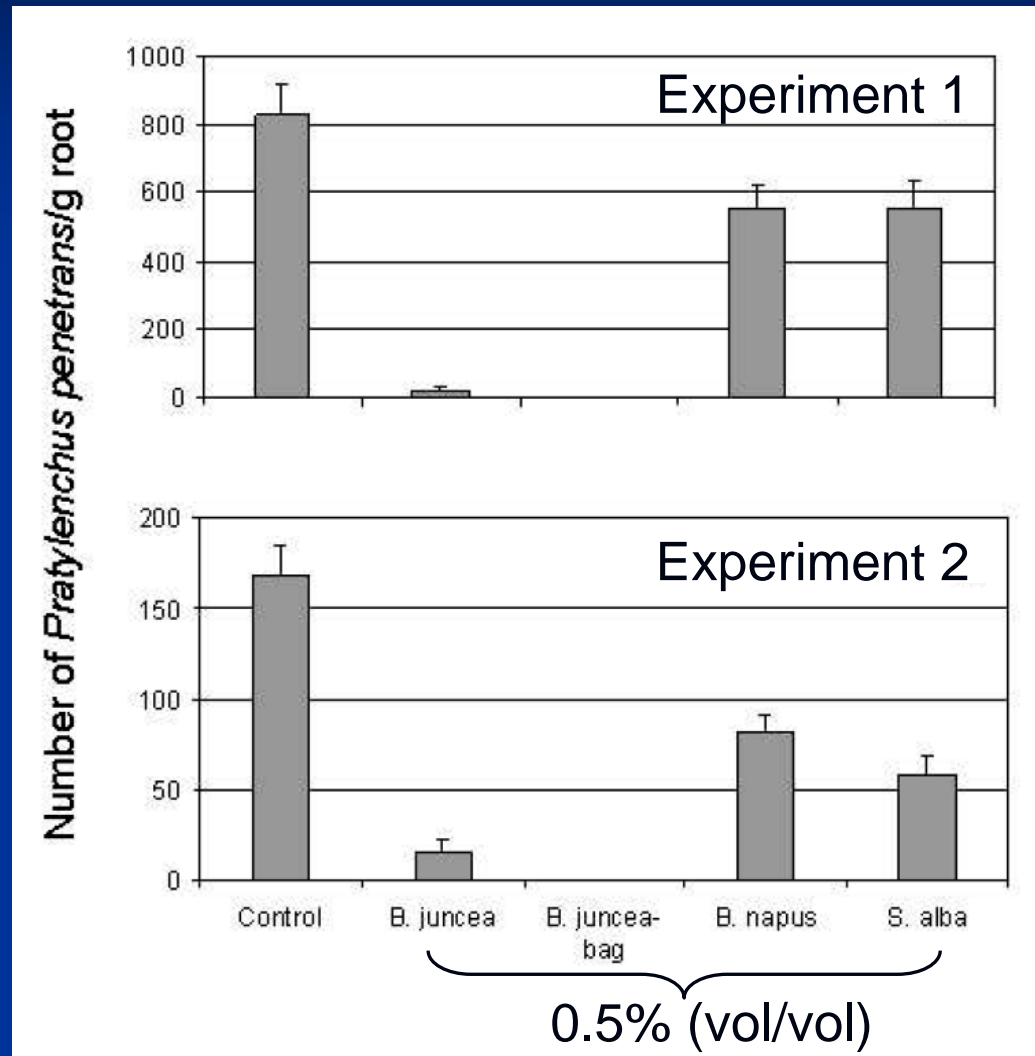
R-NH₂ \rightarrow NH₃



Seed meal nutrients

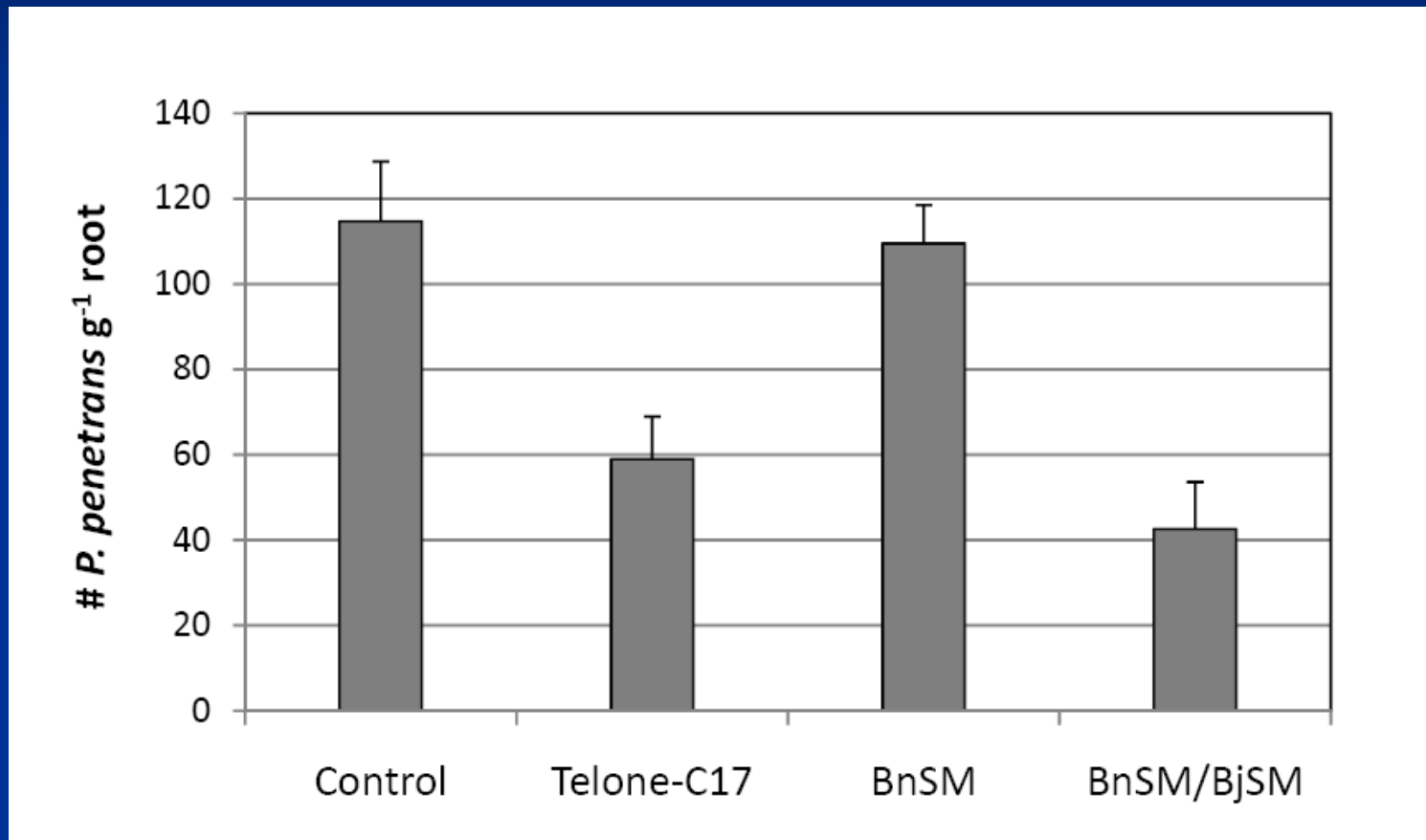
Element	Percent composition
Nitrogen	5.6 – 6.8%
Phosphorus	1.2 – 1.4%
Potassium	1.1 – 1.5%
Sulfur	0.9 – 1.6%

Superior protection conferred by *Brassica juncea* seed meal



Greenhouse
GC orchard soil
Various rootstocks

Orchard results: Control of lesion nematode by a BSM mixture

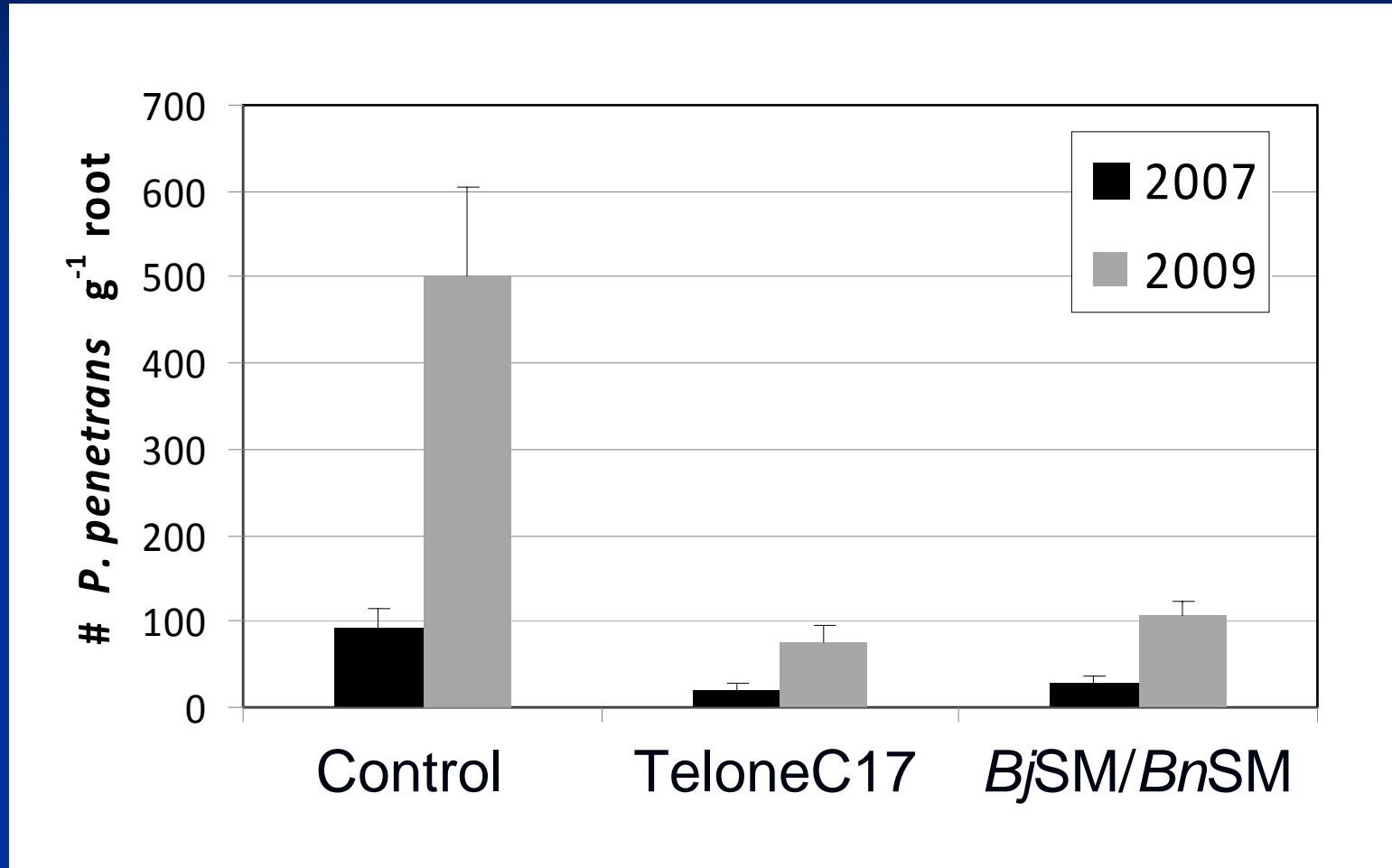


Mazzola & Brown (2009) unpublished

Commercial organic orchard

Planted May 2006 with M26 rootstock; harvested October 2006

Orchard results: Sustained control of lesion nematode



Commercial organic orchard
Planted May 2007 with Gala / M26 rootstock

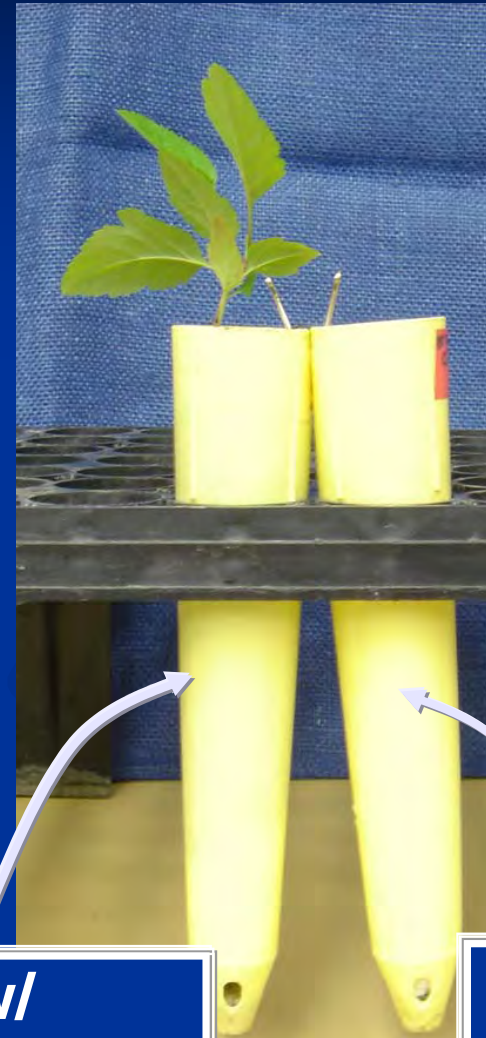
Mazzola & Brown (2009) unpublished

BSM induction of systemic plant disease resistance

Split-root assays to assess systemic resistance



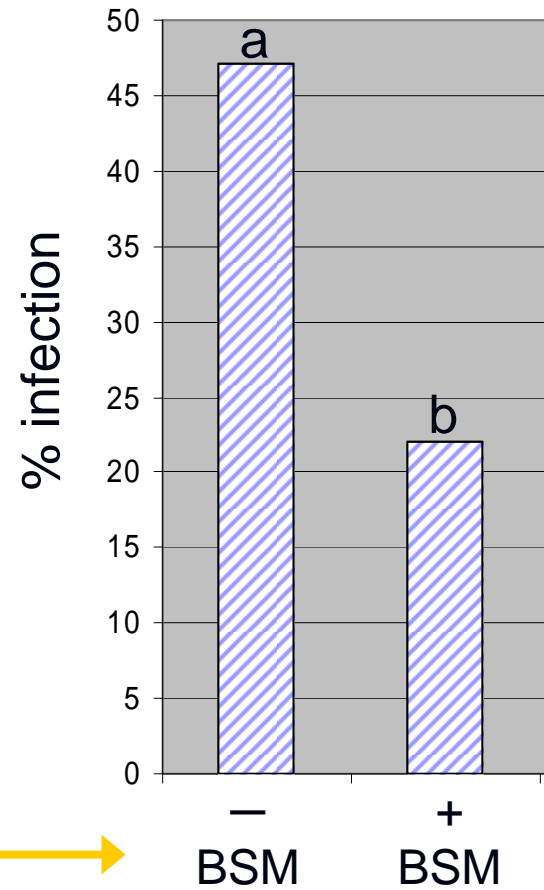
10-week old Gala seedling



w/
pathogen

w/ or w/o
additive

BSM induces **systemic** resistance against *Rhizoctonia solani* infection

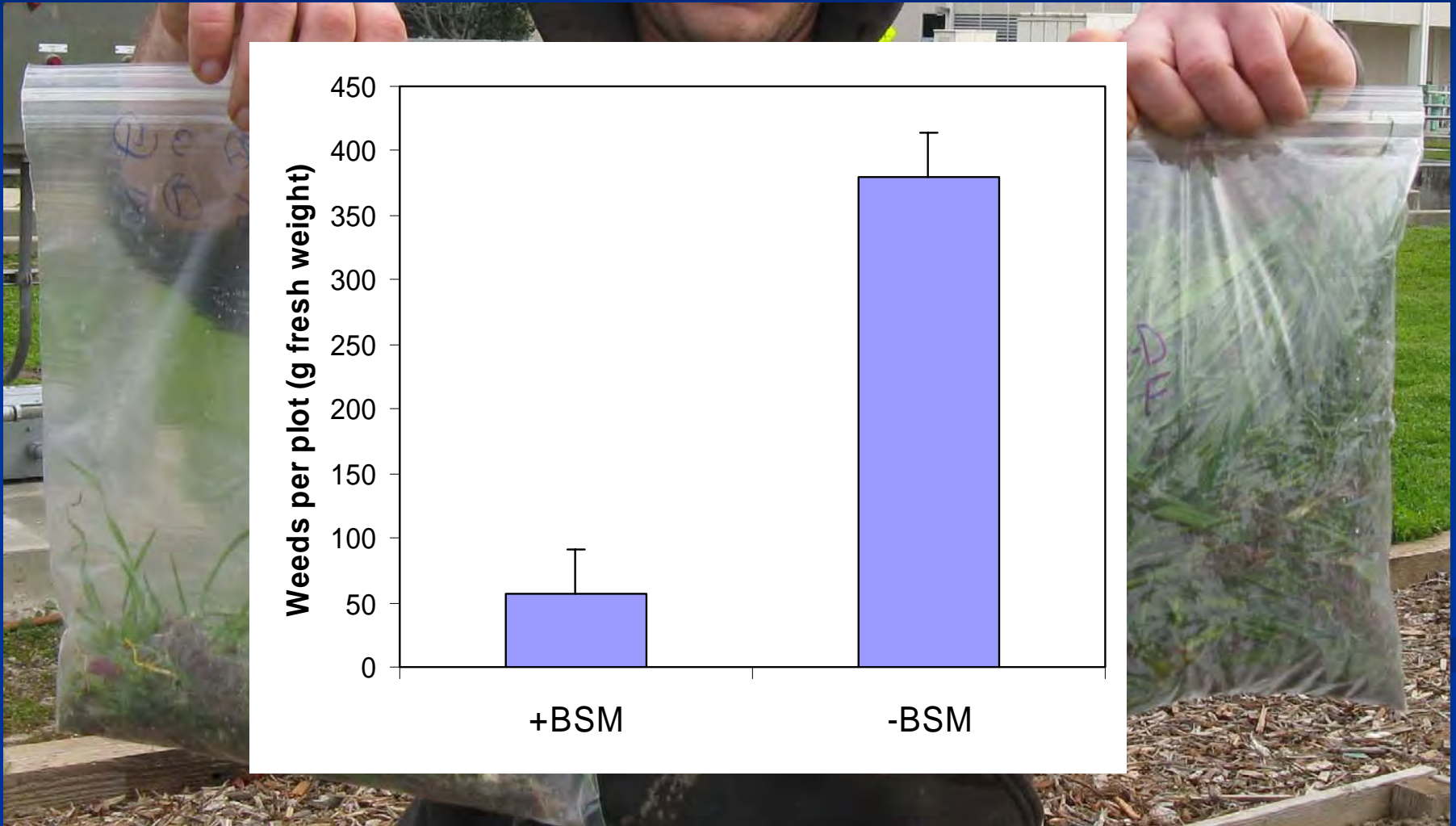
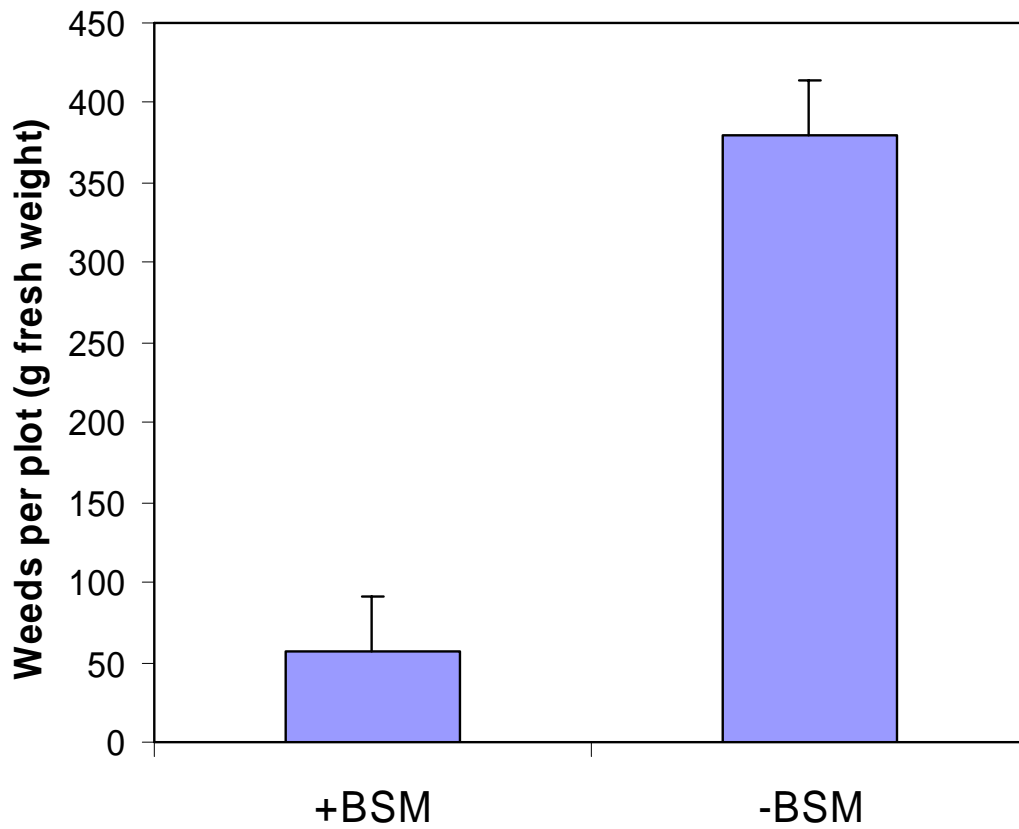


Conclusion

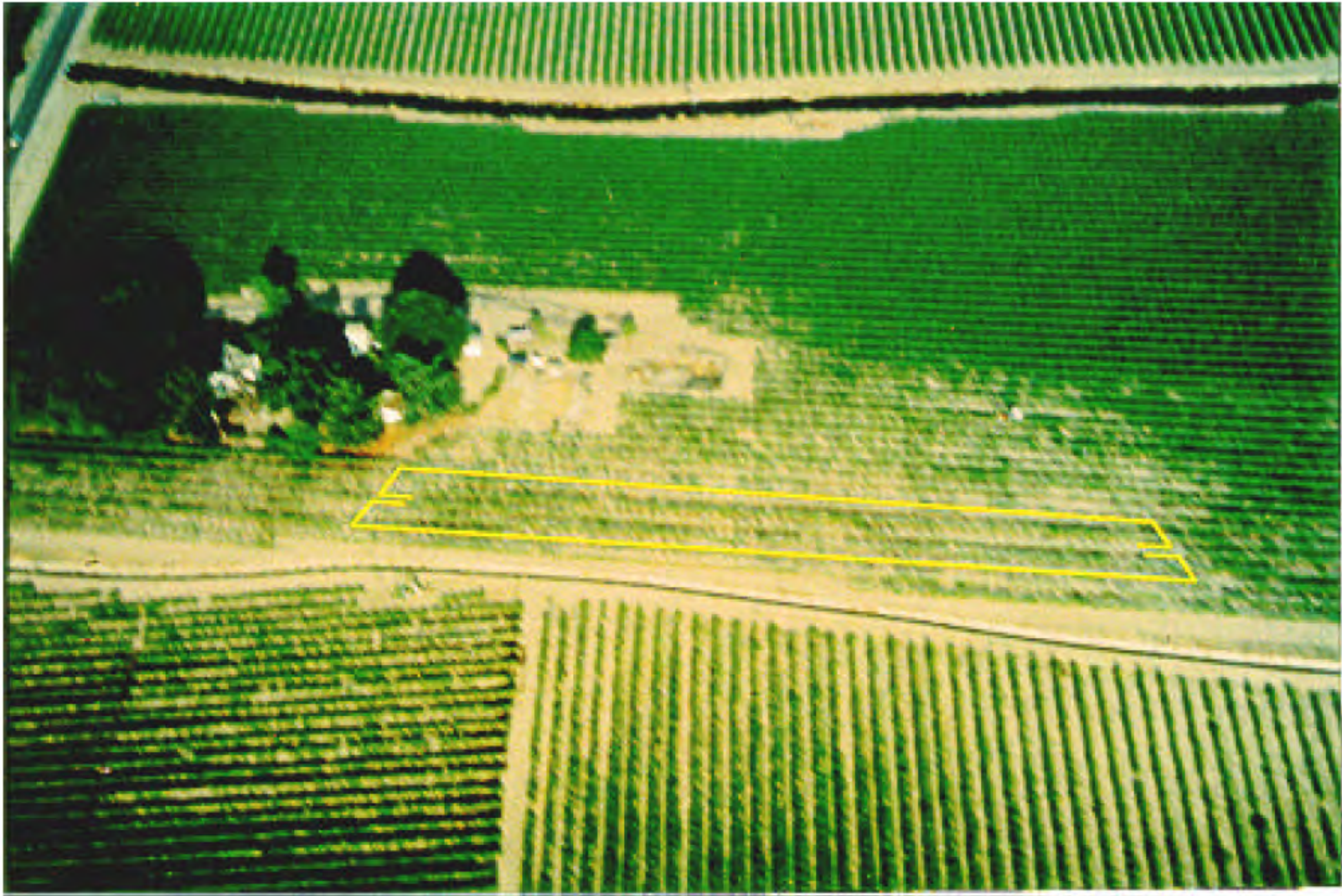
- BSM mixtures have potential application as pre-plant soil treatments for control of replant diseases.



Immediate post-application phytotoxicity of BSM controls weeds



BSM for control of vine replant disease?



Use of B_jSM on existing vineyards

Table 1. Effects of Indian mustard green manure (GM) and seed meal (SM) application in inter-row and vine row on *Meloidogyne javanica* J₂ population densities in vine row soil in vineyards (cv. Semillon), Denman, Hunter Valley, 1999–2001

Treatment	<i>M. javanica</i> J ₂ /kg soil ^A			
	Initial population density	14 WAT ^B	24 WAT	36 WAT
<i>3-year-old vineyard</i>				
Control	1540 (7.34) a	1107 (7.01) a	1790 (7.49) a	1096 (7.00) a
Inter-row GM	1540 (7.34) a	595 (6.39) a	632 (6.45) b	220 (5.39) b
Vine row GM	1495 (7.31) a	120 (4.79) c	68 (4.22) c	86 (4.46) c
Inter-row SM	1408 (7.25) a	90 (4.50) c	340 (5.83) c	276 (5.62) b
Vine row SM	1425 (7.26) a	121 (4.80) c	185 (5.22) d	76 (4.34) c
LSD ($P < 0.05$)	0.21	0.41	0.37	0.26
<i>15-year-old vineyard</i>				
Control	1670 (7.42) a	2345 (7.76) a	1465 (7.29) a	1366 (7.22) a
Inter-row GM	1152 (7.05) a	268 (5.59) bc	105 (4.65) d	880 (6.78) a
Vine row GM	1248 (7.13) a	169 (5.13) c	317 (5.76) bc	347 (5.85) b
Inter-row SM	1236 (7.12) a	165 (5.11) c	411 (6.02) b	1107 (7.01) a
Vine row SM	1188 (7.08) a	388 (5.96) b	230 (5.44) c	304 (5.72) b
LSD ($P < 0.05$)	0.19	0.68	0.60	0.28

^ABack transformed means of *M. javanica* J₂ population densities in vine row soil with transformed means ($\log e^x$) in parentheses. Within columns, means with different letters differ at $P < 0.05$ and LSD values are based on $\log e^x$ transformation of nematode population densities.

^BWAT = weeks after treatment.

Integrating vineyard pathogen
control with sustainable nutrient and
energy management

Application of composts to improve soil properties

- ↑ porosity
- ↑ water holding capacity
- ↓ bulk density
- improves plant capacity to withstand pathogens



Anaerobic digestion

- provides substrate for composting
- reduces solid waste volume
- provides energy for on-site uses



Clos du Bois



Simi

Heat, power

winery waste



Digestate

Effluent water



Irrigation



Harvested aquatic biomass

Anaerobic digestion



Feedstocks: Dairy waste

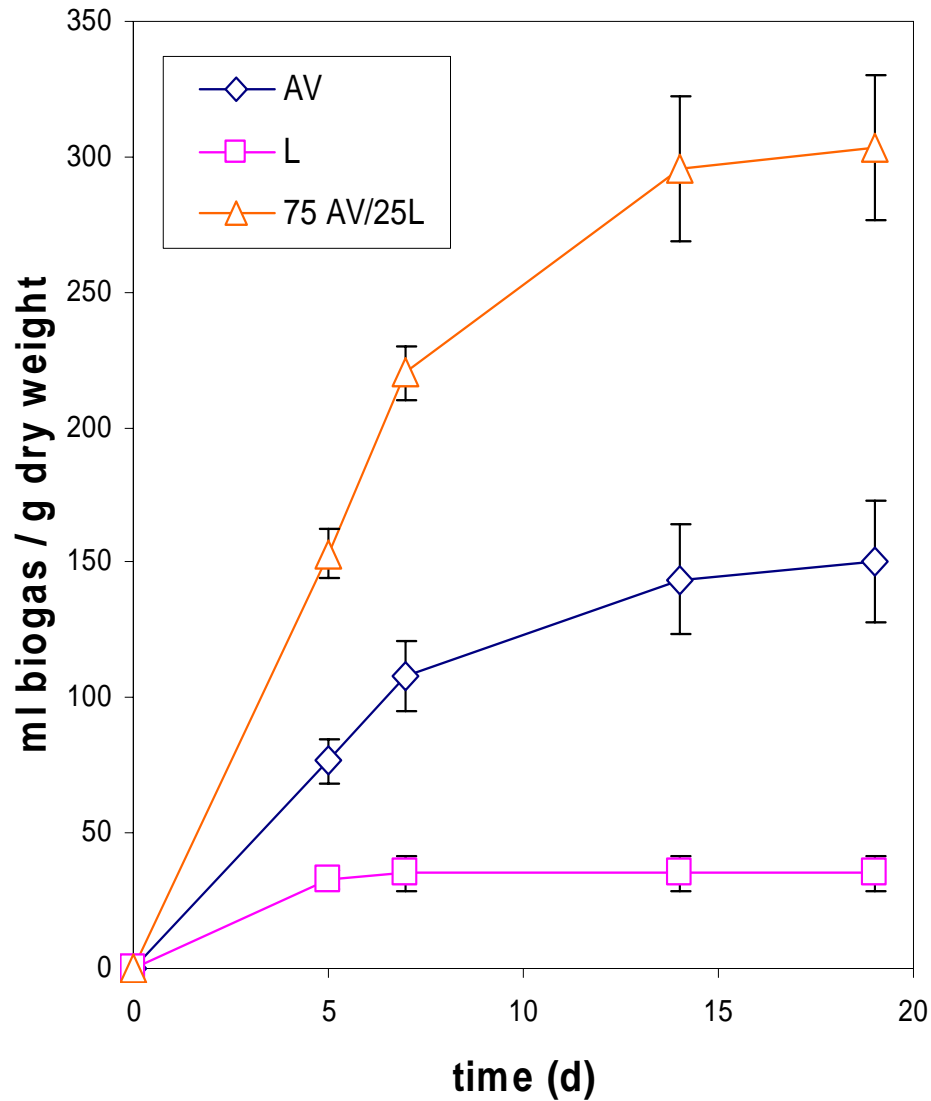
Aquatic vegetation

Winery waste

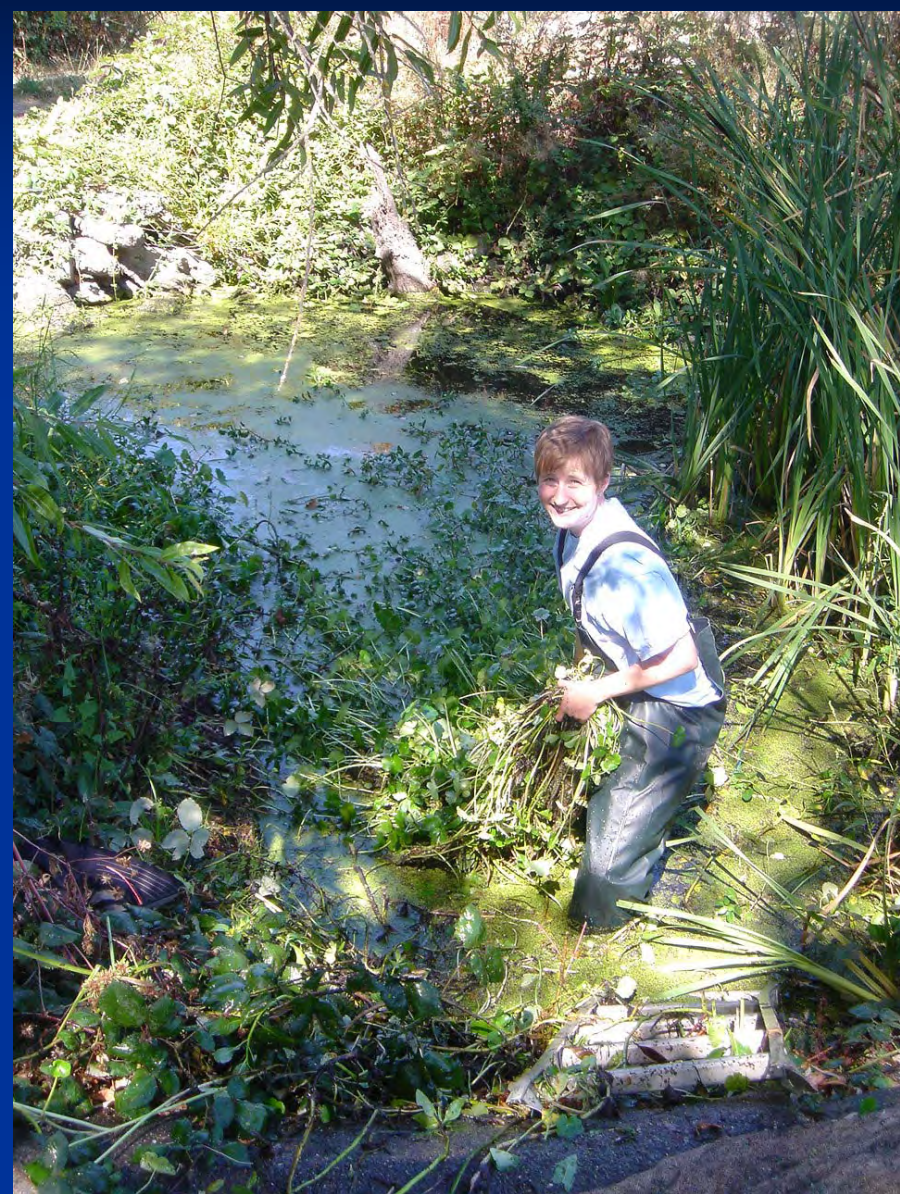
Laboratory assessment of feedstocks



Advantage of co-digestion



Aquatic vegetation: drainage channels



HANNA
WINERY & VINEYARDS

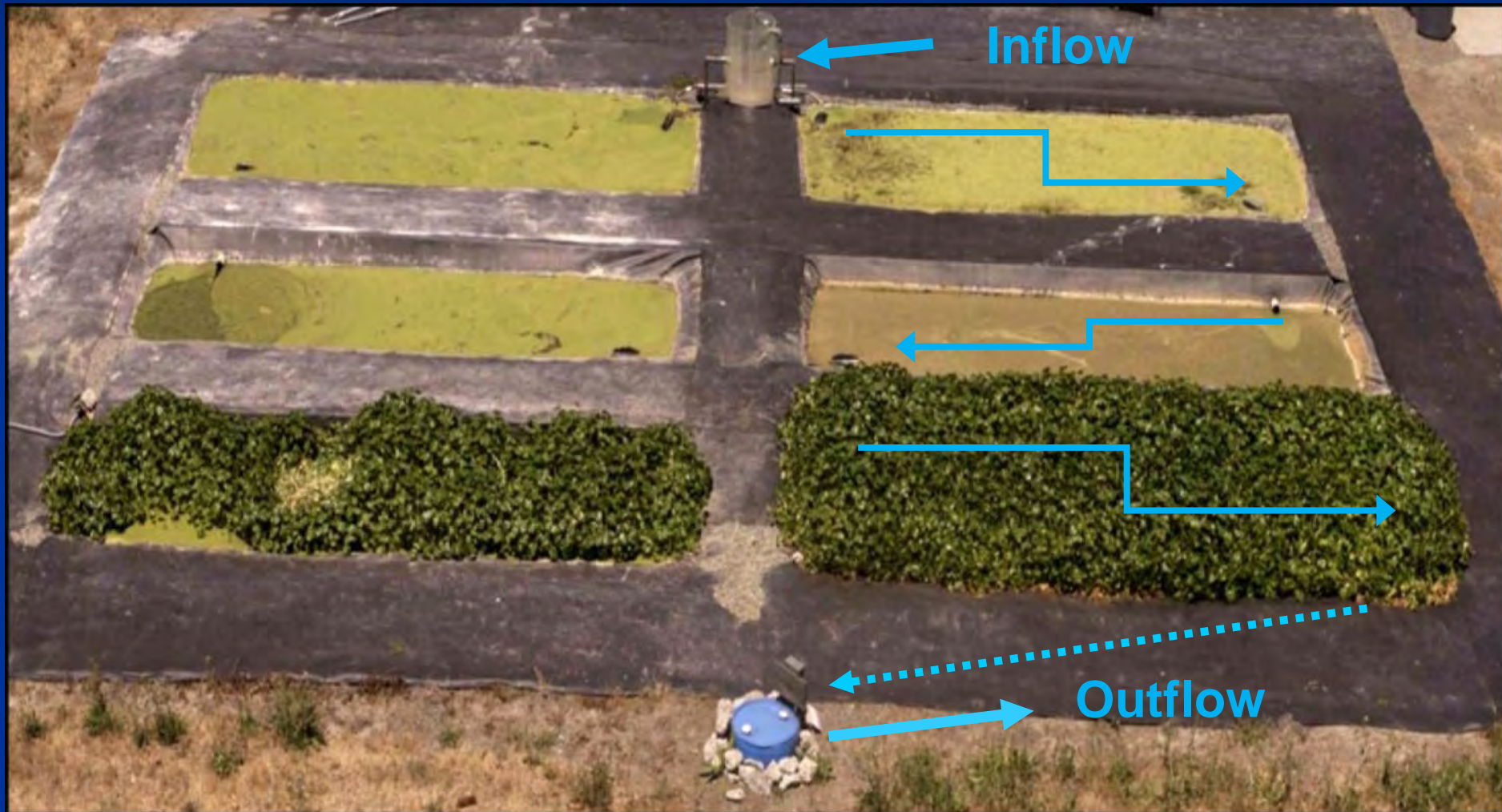


Vineyard constructed wetlands



“This year we saved 4 million gallons of water I was able to use for irrigation instead of buying it or pulling it out of the river”
-- Tim Thornhill, Parducci Winery (The Press Democrat, 1/17/2010)

Channelized Aquatic Scrubbers (CAS) at the Laguna Treatment Plant



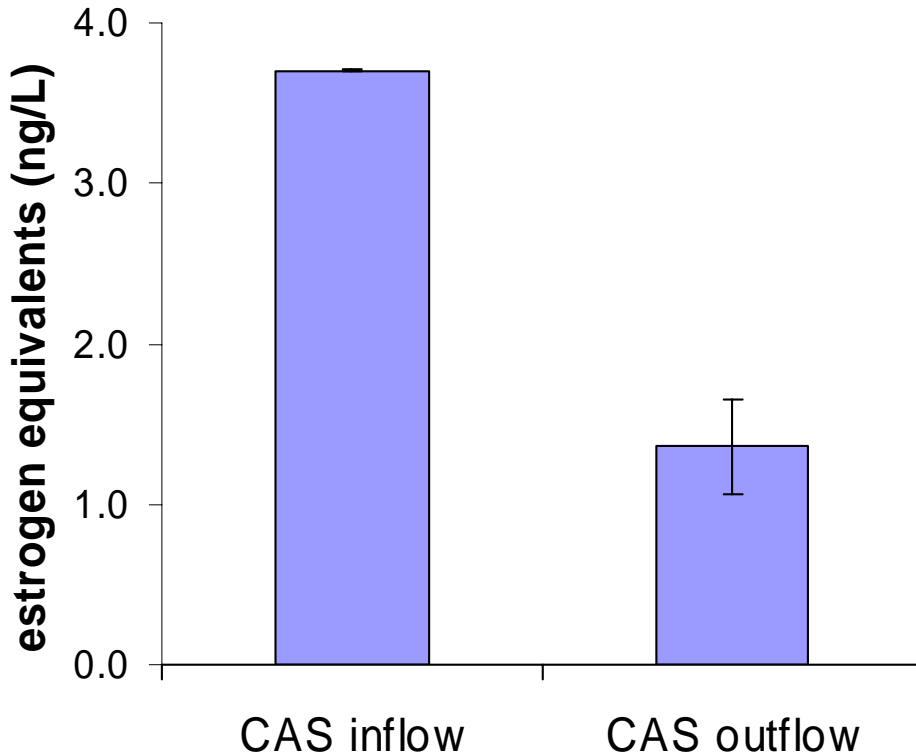
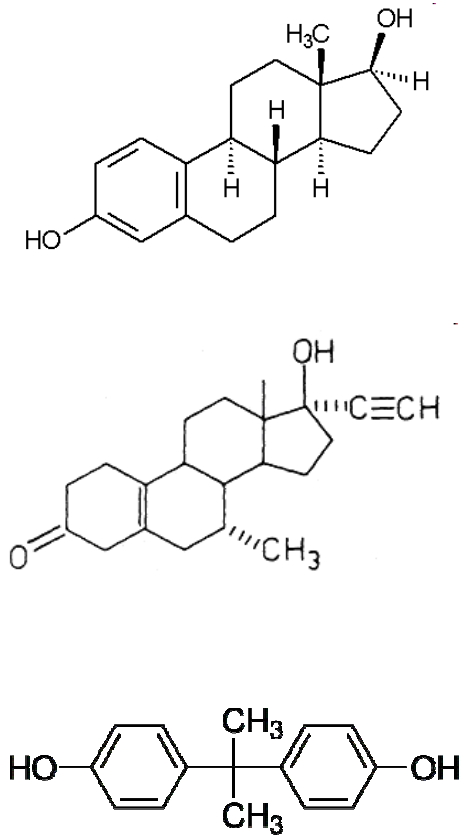
Native vegetation



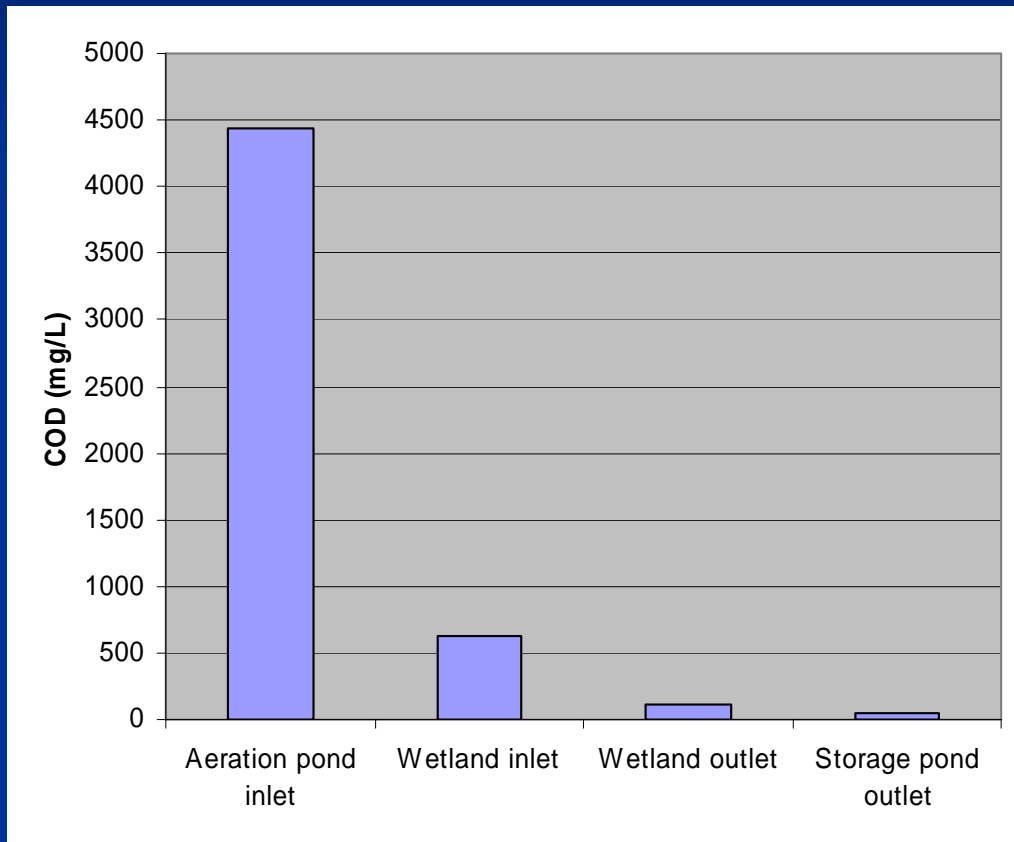
Comparative nitrate removal efficiencies

Treatment Systems	Average nitrate removal efficiency (mg N m ⁻² day ⁻¹)
Arcata Wetlands	800
Kelly Farm Wetlands	625
Prado Wetlands	522
Channelized Aquatic Scrubbers (1 July 2008 – 30 June 2009)	988

Removal of organic contaminants



COD reduction by vineyard constructed wetlands



3.3 d



Benzinger, October 2003

Heat, power

winery waste



Digestate

Effluent water



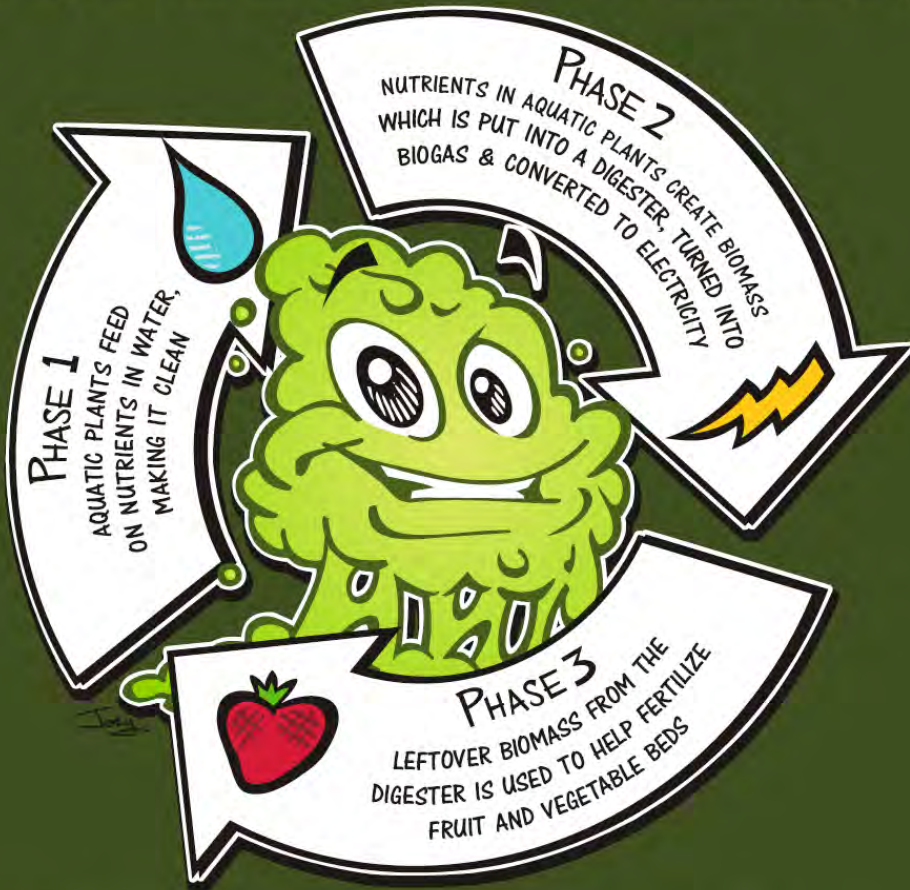
Irrigation



Harvested aquatic biomass

PRODUCING ENERGY & MORE FROM TREATED WATER!

FUEL FROM AQUATIC BIOMASS



F.A.B. Project
Laguna Subregional Treatment Plant

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